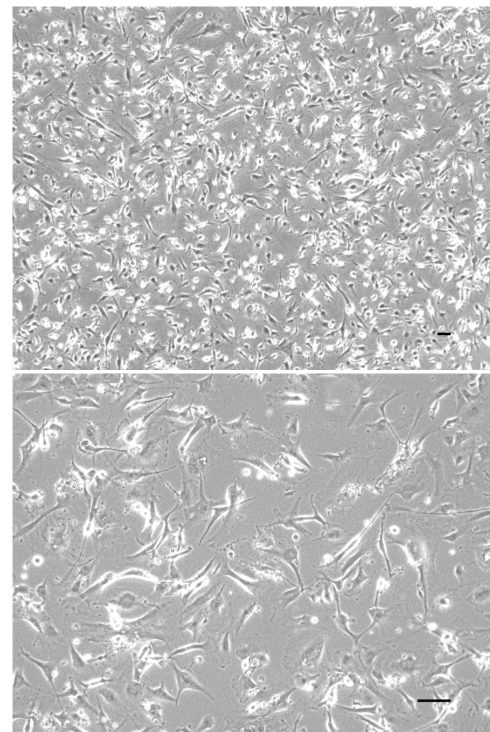
Data Sheet

Division of Rare Cancer Research,
National Cancer Center Research Institute, Japan

1． Information of the cell line

Images passage: 39



Cell line ID: **NCC-MPNST5-C1**

Organism: Human

Tissue: Soft tissue

Disease: Malignant peripheral nerve sheath t

Morphology: Fibroblast

Growth Properties: Adherent

Genetics: NF1 mutation

CDKN2A loss

Key references: PMID: 32099531

2. Culture method

Growth Medium

DMEM/F12 with fetal bovine serum 10%

DMEM/F12 GlutaMAX : Gibco, 10565042 Thermo Fisher Scientific

Fetal bovine serum; Gibco, 10270-106, Thermo Fisher Scientific

Thawing Cells

1. Thaw the vial by gentle agitation in a 37° C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium. and spin at approximately 125 x g for 5 minutes.
4. Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6). pH (7.0 to 7.6).
5. Incubate the culture at 37° C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Subculturing Procedure

Volumes are given for a 6 cm or 9 cm dish. Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes.

1. Remove and discard culture medium.
2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin0.53 mM EDTA solution to remove all traces of serum that contains trypsin inhibitor.
3. Add 1.0 to 2.0 mL of TrypsinEDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes).
Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37° C to facilitate dispersal.
4. Add 1.0 to 4.0 mL of complete growth medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37° C.

Subcultivation Ratio: 1:2 to 1:4

Medium Renewal: Every 3 to 4 days

Cryopreservation

Freeze medium: Cell Banker 1, DS-CB011, Nippon zenyaku kogyo
Storage temperature: Liquide nitrogen vapor phase

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